613 PRELIMINARY NOTES

that the metal may be removed by dialysis against dithionite plus dipyridyl or phenanthroline<sup>10,11</sup>.

We thank Miss H. Chapman for supplying the milk samples, Mr. P. Edwards for carrying out the ultracentrifuge measurements and Miss K. HARMAN and Miss M. McGartoll for technical assistance. The work was supported by grants to the Chester Beatty Research Institute (Institute of Cancer Research: Royal Cancer Hospital) from the Medical Research Council, the British Empire Cancer Campaign for Research and by the Public Health Service Research Grant No. CA-03188-10 from the National Cancer Institute, U.S. Public Health Service.

Chester Beatty Research Institute, Institute of Cancer Research: Royal Cancer Hospital, London (Great Britain)

L. I. HART R. C. Bray

- I R. C. Bray, in P. D. Boyer, H. Lardy and K. Myrbäck, The Enzymes, Vol. 7, Academic Press, New York, 1963, p. 533.
- 2 R. C. Bray, R. Pettersson and A. Ehrenberg, Biochem. J., 81 (1961) 178.
- 3 G. PALMER, R. C. BRAY AND H. BEINERT, J. Biol. Chem., 239 (1964) 2657.

- 4 F. Bergel and R. C. Bray, Biochem. J., 73 (1958) 182.
  5 P. G. Avis, F. Bergel and R. C. Bray, J. Chem. Soc., (1955) 1100.
  6 R. C. Bray, A. J. Chisholm, L. I. Hart, L. S. Meriwether and D. C. Watts, in E. C. SLATER, Flavins and Flavoproteins (BBA Library, Vol. 8), Elsevier, Amsterdam, 1966, p. 117.
- 7 H. B. BURCH, in S. P. COLOWICK AND N. O. KAPLAN, Methods in Enzymology, Academic Press, New York, 1957, p. 960.
- 8 P. G. Avis, F. Bergel and R. C. Bray, *J. Chem. Soc.*, (1956) 1219. 9 C. E. Johnson, P. F. Knowles and R. C. Bray, *Biochem. J.*, 103 (1967) 10C.
- 10 E. BAYER AND W. VOELTER, Biochem. Biophys. Acta, 113 (1966) 632.
- 11 M. UOZUMI, R. HAYASHIKAWA AND L. H. PIETTE, Arch. Biochem. Biophys., 119 (1967) 288.
- 12 P. Andrews, R. C. Bray, P. Edwards and K. V. Shooter, Biochem. J., 93 (1964) 627.

Received July 27th, 1967

Biochim. Biophys. Acta, 146 (1967) 611-613

BBA 61147

## Preparation of an adrenal steroid 21-hydroxylating system exhibiting activity in the absence of the carbon monoxide-binding pigment, cytochrome P-450

Cytochrome P-450 has been implicated as the oxygen-activating component in adrenal steroid hydroxylation using various mitochondrial and microsomal preparations<sup>1-3</sup>. We have been unable to confirm the necessity for the presence of cytochrome P-450 in active, purified preparations of steroid 21-hydroxylase (EC 1.14.1.8).

Sheep adrenal glands were processed, fractionated and assayed for 21-hydroxylating activity, using 17-hydroxyprogesterone as substrate, as previously described4. The most active dialyzed ammonium sulfate precipitate (0.3 S) was frozen and thawed once, and centrifuged for 20 min at 10 000 × g. About 50% of the activity remained in the supernatant, which was further fractionated by gel filtration on a column (5 cm  $\times$  100 cm) of Sephadex G-200 at 4°. The column was equilibrated with o.I M phosphate buffer (pH 6.8) containing reduced glutathione, I mg/ml, and

614 PRELIMINARY NOTES

eluted with the same buffer with an upward flow rate of 20 ml/h. The applied material was resolved into five components of which only the first fraction, traveling with the the front, exhibited hydroxylating activity. Several different homogenates which had been stored for various lengths of time up to 6 months at -20° were processed in this way. Some of these active preparations were concentrated by ultrafiltration to yield protein concentrations of from 1.88 to 11.5 mg/ml (ref. 5). These preparations were capable of 21-hydroxylating 17-hydroxyprogesterone to 17,21-dihydroxy-4pregnene-3,20-dione with an average production of 58 mµmoles/mg protein per h. These active preparations were analyzed for the presence of the carbon monoxidebinding pigment, cytochrome P-450, by the methods of ESTABROOK, COOPER AND ROSENTHAL<sup>1</sup> and OMURA AND SATO<sup>6</sup>. The difference spectra were obtained with a Bausch and Lomb 505 recording spectrophotometer. There was no evidence for the presence of cytochrome P-450 in any of these active preparations regardless of protein concentration. Identical results were obtained with preparations stored as such, or in the presence of 50% glycerol, at  $-20^{\circ}$ . The absence of P-450 in active preparations obtained by fractionation in the presence of reduced glutathione and subsequent storage in glycerol is all the more pertinent since ICHIKAWA AND YAMANO7 recently reported the reversal of P-420 to P-450 by treatment with reduced glutathione and the stabilizing influence of glycerol on P 450.

This work was supported by Grant AM 03269 08, National Institute of Arthritis and Metabolic Diseases, National Institutes of Health, U.S. Public Health Service.

Department of Endocrinology, Southwest Foundation for Research and Education, San Antonio, Texas (U.S.A.) Charles Matthijssen Joyce E. Mandel

I R. W. ESTABROOK, D. Y. COOPER AND O. ROSENTHAL, Biochem. Z., 338 (1963) 741.

- 2 R. W. ESTABROOK, J. B. SCHENKMAN, W. CAMMER, H. REMMER, D. Y. COOPER, S. NARASIM-HULU AND O. ROSENTHAL, in K. BLOCH AND O. HAYAISHI, Biological and Chemical Aspects of Oxygenases, Maruzen, Tokyo, 1966, p. 153.
- 3 L. D. Wilson, D. H. Nelson and B. W. Harding, Biochim. Biophys. Acta, 99 (1965) 391.
- 4 C. MATTHIJSSEN AND J. E. MANDEL, Biochim. Biophys. Acta, 82 (1964) 138.
- 5 O. WARBURG AND W. CHRISTIAN, Biochem. Z., 310 (1941) 384.
- 6 T. OMURA AND R. SATO, J. Biol. Chem., 239 (1964) 2370.
- 7 Y. ICHIKAWA AND T. YAMANO, Biochim. Biophys. Acta, 131 (1967) 490.

Received August 7th, 1967

Biochim. Biophys. Acta, 146 (1967) 613-614